

Phenylpropanoid Esters of Rhamnose from *Buddleja asiatica*

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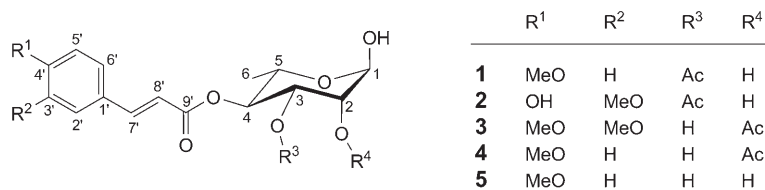
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Four new phenylpropanoid esters of rhamnose, asiatisides A–D, along with the known compounds, buergeriside C₁ (**5**), *p*-methoxycinnamic acid, ferulic acid, and *O*-methylferulic acid were obtained from the aerial parts of *Buddleja asiatica* LOUR by chromatographic methods. The new compounds were elucidated as 3-*O*-acetyl-4-*O*-(*p*-methoxycinnamoyl)- α -L-rhamnopyranose (**1**), 3-*O*-acetyl-4-*O*-feruloyl- α -L-rhamnopyranose (**2**), 2-*O*-acetyl-4-*O*-(*O*-methylferuloyl)- α -L-rhamnopyranose (**3**), 2-*O*-acetyl-4-*O*-(*p*-methoxycinnamoyl)- α -L-rhamnopyranose (**4**) by spectral data (1D-, 2D-NMR, and MS), respectively.

Introduction. – The genus *Buddleja* belonging to the family Buddlejaceae consists of shrubs or half-shrubs, distributed in tropical and subtropical regions [1]. The flowers, leaves, and roots of various species of *Buddleja* are used in traditional medicine in several countries [2]. In particular, the leaves and flowers of *B. globosa* are used in Chile for washing wounds and treating ulcers [2]. ‘*Mi-meng-hua*’, prepared from the flowers of *B. officinalis*, is a traditional Chinese medicine used for the treatment of conjunctival congestion and clustered nebulae [3], and *B. asiatica* is named ‘*Qi-li-xiang*’ by local people in Guangxi province. Its roots, stems, and leaves are used as a popular traditional Chinese medicine for the treatment of fever, ache, diarrhea, and articular rheumatism [4]. Previous studies on *B. officinalis* and *B. globosa* led to the isolation of triterpenoid glycosides [5a][5b], flavonoid triglycosides [6], iridoid glycosides [7], sesquiterpenes [8], diterpenoids [9], and saponins [10]. Up to now, few chemical constituents [11a – c] have been reported from *B. asiatica*, including buddlin, flavonoids, and sterols. In the present study, we report the isolation and structure determination of four new phenylpropanoid esters of rhamnose named asiatisides A–D (Fig. 1), along with four known compounds from aerial parts of *B. asiatica* LOUR.

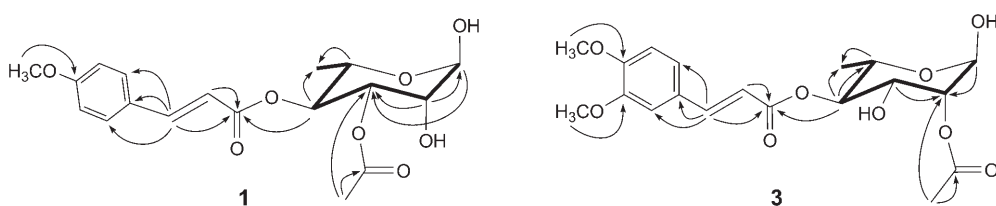
Results and Discussion. – Compound **1** was obtained as a white amorphous powder ($[\alpha]_{\text{D}}^{22} = +85.7$ ($c = 0.21$, acetone)). The molecular formula C₁₈H₂₂O₈ was determined by the HR-ESI-MS spectrum (m/z 389.1211 ($[M + \text{Na}]^+$; calc. 389.1212)) in combination with ¹H-, ¹³C-NMR, and DEPT spectra, corresponding to eight degrees of unsaturation. The ¹H-, ¹³C-NMR, and DEPT spectra displayed an α -L-rhamnosyl unit [12] on the basis of a H-atom signal at $\delta(\text{H})$ 5.12 (*dd*, $J = 1.8, 4.0$, H–C(1)¹), a secondary Me group signal at $\delta(\text{H})$ 1.13 (*d*, $J = 6.3$, Me(6)) as well as C-atom signals for

¹⁾ Arbitrary numbering; for systematic names, see *Exper. Part*.

Fig. 1. Chemical structures of **1–5**¹⁾

the anomeric C-atom ($\delta(\text{C})$ 95.3 (*d*)), a Me group ($\delta(\text{C})$ 18.1 (*q*)), and other four CH groups ($\delta(\text{C})$ 72.7 (*d*), 72.0 (*d*), 70.6 (*d*), 66.9 (*d*)). The identification of the sugar unit was continued by hydrolysis with 10% HCl to afford L-rhamnose, which was confirmed by TLC comparison with an authentic sample and determination of its optical rotation value ($[\alpha]_{\text{D}}^{18} = +12.2$; MeOH) [13].

In addition to the above described signals, the ¹H-NMR spectrum of **1** exhibited signals of a *p*-substituted cinnamoyl moiety and a MeO group [14a][14b]: four H-atoms of an aromatic ring (*AABB* system, $\delta(\text{H})$ 7.65 (*d*, $J = 8.7$, H–C(2'), H–C(6')), and 6.98 (*d*, $J = 8.7$, H–C(3'), H–C(5')), as well as two (*E*)-olefinic H-atoms (*AB* system, $\delta(\text{H})$ 6.38, 7.64 (*d*, $J = 16.1$, 2×1 H)). The ¹³C-NMR spectrum also exhibited signals of a cinnamoyl moiety, *i.e.*, six aromatic C-atoms ($\delta(\text{C})$ 162.6, 130.9, 130.9, 127.9, 115.2, 115.2) and two olefinic C-atoms ($\delta(\text{C})$ 115.8, 145.7). In the HMBC spectrum (Fig. 2), the MeO signal at $\delta(\text{H})$ 3.84 (*s*) showed a cross-peak with the signal at $\delta(\text{C})$ 162.6 (*s*, C(4')), indicating a MeO substituent at C(4'). In the ¹H,¹H-COSY spectrum (Fig. 2), a signal at $\delta(\text{H})$ 4.10–4.12 (*m*, 1 H) showed correlations with Me(6) ($\delta(\text{H})$ 1.13), assigning the signal to H–C(5). Likewise, the signal at $\delta(\text{H})$ 4.05–4.06 (*m*, 1 H) correlated with H–C(1) ($\delta(\text{H})$ 5.12), was attributed to H–C(2). Two remaining overlapping signals at $\delta(\text{H})$ 5.27–5.29 (*m*, 2 H) were assigned to H–C(3) and H–C(4). Further, these two H-atoms corresponded to $\delta(\text{C})$ 72.7 (*d*) and 72.0 (*d*) in the HSQC spectrum. Considering that $\delta(\text{C})$ 72.7 (*d*) showed a correlation with H–C(1) in the HMBC spectrum, this signal was attributed to C(3). The other signal ($\delta(\text{C})$ 72.0, *d*) was assigned to C(4), which could be supported by a correlation of the signal of H–C(6) with $\delta(\text{C})$ 72.0 (*d*). In the HMBC experiment (Fig. 2), two overlapping H-atom signals at $\delta(\text{H})$ 5.27–5.29 (*m*, 2 H) were correlated with the CO group at $\delta(\text{C})$ 166.7 (C(9'), *s*) and an Ac CO group at $\delta(\text{C})$ 170.5 (*s*), suggesting that the cinnamoyl moiety and the AcO group might be connected to C(4) and C(3), respectively. The respective position of the acyl residues was deduced from the correlation of the Ac

Fig. 2. Key COSY (—) and HMBC (---) correlations for **1** and **3**

H-atoms ($\delta(\text{H})$ 1.93 (s, 3 H)) and C(3). Thus, compound **1** was identified as 3-*O*-acetyl-4-*O*-(*p*-methoxycinnamoyl)- α -L-rhamnopyranose¹), and named asiatiside A. Full assignments of the signals of ¹H- and ¹³C-NMR of **1** were determined by detailed analysis of ¹H, ¹H-COSY, HSQC, and HMBC spectra (Tables 1 and 2).

Table 1. ¹H-NMR Data of **1–4**¹). At 500 MHz in (D₆)acetone; δ in ppm, *J* in Hz.

Position	1	2	3	4
1	5.12 (<i>dd</i> , <i>J</i> = 1.8, 4.0)	5.10 (<i>dd</i> , <i>J</i> = 1.7, 4.1)	5.07–5.09 (<i>m</i>) ^a	5.06–5.08 (<i>m</i>) ^a
2	4.05–4.06 (<i>m</i>)	4.04–4.05 (<i>m</i>)	5.07–5.09 (<i>m</i>) ^a	5.06–5.08 (<i>m</i>) ^a
3	5.27–5.29 (<i>m</i>) ^a	5.26–5.29 (<i>m</i>) ^a	4.14–4.16 (<i>m</i>)	4.13–4.17 (<i>m</i>)
4	5.27–5.29 (<i>m</i>) ^a	5.26–5.29 (<i>m</i>) ^a	4.99 (br. <i>t</i> , <i>J</i> = 9.8)	4.99 (br. <i>t</i> , <i>J</i> = 9.8)
5	4.10–4.12 (<i>m</i>)	4.09–4.11 (<i>m</i>)	4.03–4.06 (<i>m</i>)	4.03–4.07 (<i>m</i>)
6	1.13 (<i>d</i> , <i>J</i> = 6.3)	1.11 (<i>d</i> , <i>J</i> = 6.3)	1.11 (<i>d</i> , <i>J</i> = 6.5)	1.13 (<i>d</i> , <i>J</i> = 6.5)
2'	7.65 (<i>d</i> , <i>J</i> = 8.7)	7.18 (<i>d</i> , <i>J</i> = 2.0)	7.34 (<i>d</i> , <i>J</i> = 1.8)	7.63 (<i>d</i> , <i>J</i> = 8.7)
3'	6.98 (<i>d</i> , <i>J</i> = 8.7)	–	–	6.99 (<i>d</i> , <i>J</i> = 8.7)
5'	6.98 (<i>d</i> , <i>J</i> = 8.7)	6.98 (<i>d</i> , <i>J</i> = 8.3)	6.98 (<i>d</i> , <i>J</i> = 8.5)	6.99 (<i>d</i> , <i>J</i> = 8.7)
6'	7.65 (<i>d</i> , <i>J</i> = 8.7)	7.13 (<i>dd</i> , <i>J</i> = 2.0, 8.3)	7.21 (<i>dd</i> , <i>J</i> = 1.8, 8.5)	7.63 (<i>d</i> , <i>J</i> = 8.7)
7'	7.64 (<i>d</i> , <i>J</i> = 16.1)	7.58 (<i>d</i> , <i>J</i> = 15.9)	7.64 (<i>d</i> , <i>J</i> = 16.0)	7.67 (<i>d</i> , <i>J</i> = 16.0)
8'	6.38 (<i>d</i> , <i>J</i> = 16.1)	6.32 (<i>d</i> , <i>J</i> = 15.9)	6.45 (<i>d</i> , <i>J</i> = 16.0)	6.42 (<i>d</i> , <i>J</i> = 16.0)
MeO–C(3')		3.88 (<i>s</i>)	3.84 (<i>s</i>)	
MeO–C(4')	3.84 (<i>s</i>)		3.88 (<i>s</i>)	3.85 (<i>s</i>)
Ac	1.93 (<i>s</i>)	1.92 (<i>s</i>)	2.09 (<i>s</i>)	2.09 (<i>s</i>)

^a) Overlapping signals.

Table 2. ¹³C-NMR Data of **1–4**¹). At 125 MHz in (D₆)acetone, δ in ppm. Assignments are based on DEPT, HSQC, and HMBC spectra.

Position	1	2	3	4
1	95.3 (<i>d</i>)	95.4 (<i>d</i>)	92.6 (<i>d</i>)	92.6 (<i>d</i>)
2	70.6 (<i>d</i>)	70.6 (<i>d</i>)	74.4 (<i>d</i>)	74.4 (<i>d</i>)
3	72.7 (<i>d</i>)	72.7 (<i>d</i>)	67.9 (<i>d</i>)	67.9 (<i>d</i>)
4	72.0 (<i>d</i>)	72.0 (<i>d</i>)	75.3 (<i>d</i>)	75.3 (<i>d</i>)
5	66.9 (<i>d</i>)	66.9 (<i>d</i>)	66.7 (<i>d</i>)	66.7 (<i>d</i>)
6	18.1 (<i>q</i>)	18.1 (<i>q</i>)	18.0 (<i>q</i>)	18.0 (<i>q</i>)
1'	127.9 (<i>s</i>)	128.5 (<i>s</i>)	128.1 (<i>s</i>)	127.9 (<i>s</i>)
2'	130.9 (<i>d</i>)	112.3 (<i>d</i>)	110.9 (<i>d</i>)	130.8 (<i>d</i>)
3'	115.2 (<i>d</i>)	147.8 (<i>s</i>)	152.6 (<i>s</i>)	115.2 (<i>d</i>)
4'	162.6 (<i>s</i>)	150.8 (<i>s</i>)	150.6 (<i>s</i>)	163.5 (<i>s</i>)
5'	115.2 (<i>d</i>)	114.6 (<i>d</i>)	112.3 (<i>d</i>)	115.2 (<i>d</i>)
6'	130.9 (<i>d</i>)	122.5 (<i>d</i>)	123.7 (<i>d</i>)	130.8 (<i>d</i>)
7'	145.7 (<i>d</i>)	146.1 (<i>d</i>)	145.9 (<i>d</i>)	145.4 (<i>d</i>)
8'	115.8 (<i>d</i>)	115.9 (<i>d</i>)	116.3 (<i>d</i>)	116.2 (<i>d</i>)
9'	166.7 (<i>s</i>)	166.7 (<i>s</i>)	167.1 (<i>s</i>)	167.1 (<i>s</i>)
MeO–C(3')		56.3 (<i>q</i>)	56.0 (<i>q</i>)	
MeO–C(4')	55.8 (<i>q</i>)		56.0 (<i>q</i>)	55.7 (<i>q</i>)
MeCO	170.5 (<i>s</i>)	170.5 (<i>s</i>)	170.8 (<i>s</i>)	170.8 (<i>s</i>)
MeCO	20.9 (<i>q</i>)	20.9 (<i>q</i>)	20.9 (<i>q</i>)	20.9 (<i>q</i>)

The molecular formula of **2** was determined as $C_{18}H_{22}O_9$ by the HR-ESI mass spectrum (m/z 405.1161 ($[M + Na]^+$; calc. 405.1161)). The 1H -, ^{13}C -NMR, and DEPT spectra of **2** displayed similarities to those of **1**, except for one more OH group in the cinnamoyl moiety of **2**. In the 1H -NMR spectrum of **2**, ABX system signals of three H-atoms of an aromatic ring ($\delta(H)$ 7.18 ($d, J = 2.0$), 7.13 ($dd, J = 2.0, 8.3$), and 6.98 ($d, J = 8.3$)) gave a feruloyl moiety [14b][15], which was also confirmed by the HMBC spectrum. Thus, compound **2** was identified as 3-*O*-acetyl-4-*O*-feruloyl- α -L-rhamnopyranose¹) and named asiatiside B.

Compound **3** was obtained as a white amorphous powder. The molecular formula of **3** was determined as $C_{19}H_{24}O_9$ by the HR-ESI mass spectrum (m/z 419.1319 ($[M + Na]^+$; calc. 419.1318)). Comparison of the 1H -, ^{13}C -NMR, and DEPT spectra of **3** with those of **2** displayed similarities of **2** and **3**, except for the chemical shifts of the sugar unit and one more MeO group. As for compound **1**, the signals for Me(6) ($\delta(H)$ 1.11, d) and for three H-atoms (4.03–4.06 (m , H–C(5)), 4.99 (br. t , H–C(4)), 4.14–4.16 (m , H–C(3))) were assigned to the sugar moiety by careful analysis of the 1H , 1H -COSY spectrum. Two remaining overlapping signals at $\delta(H)$ 5.07–5.09 (m , 2 H) were assigned to H–C(2) and H–C(1). In the HSQC spectrum, these two H-atoms were correlated to $\delta(C)$ 74.4 (d) and 92.6 (d), respectively. In the HMBC spectrum (Fig. 2), the Ac Me group at $\delta(H)$ 2.09 (s , 3 H) showed a cross-peak with C(2) ($\delta(C)$ 74.4, d), indicating that the Ac group was connected to C(2). Furthermore, H–C(4) ($\delta(H)$ 4.99) was correlated with the CO group at $\delta(C)$ 167.1 (s , C(9')), suggesting that the feruloyl moiety was located at C(4). In addition, the MeO signals at $\delta(H)$ 3.88 (s) and 3.84 (s) were correlated with the C-atoms $\delta(C)$ 150.6 and 152.6, placing the two MeO groups at C(4') and C(3'), respectively. Therefore, compound **3** was identified as 2-*O*-acetyl-4-*O*-(*O*-methylferuloyl)- α -L-rhamnopyranose¹) and named asiatiside C.

Compound **4** has the molecular formula $C_{18}H_{22}O_8$ as evidenced by the HR-ESI mass spectrum (m/z 389.1203 ($[M + Na]^+$; calc. 389.1212)). Compounds **4** and **1** have the same molecular formula. The 1H -, ^{13}C -NMR, and DEPT spectra of **4** displayed similarities to those of **1**, except for the chemical shifts of the sugar unit. Comparison of the 1H -, ^{13}C -NMR, and DEPT spectra of **4** with those of **3** showed that there are the similar chemical shifts for the sugar unit, indicating the same substituents in the sugar unit. Therefore, compound **4** was identified as 2-*O*-acetyl-4-*O*-(*p*-methoxycinnamoyl)- α -L-rhamnopyranose¹) and named asiatiside D.

In the 1H -NMR spectra of **1–4**, a second set of signals of weaker intensity was assigned to the corresponding β -L-rhamnose derivatives.

In previous studies, phenylpropanoid esters of rhamnose display quite interesting pharmacological properties including antioxidant, and antiviral activities, as well as inhibition of blood platelet aggregation and LTB_4 synthesis [16]. Many phenylpropanoid esters of rhamnose have been isolated from plants of the genus *Scrophularia* (Scrophulariaceae), such as ningposides A–D [14b][17] and buergerisides A₁, B₁, B₂, and C₁ [18]. In these known compounds, the phenylpropanoid moiety is usually located at C(2) or C(3) of the sugar unit. However, in our study, this substituent was attached to C(4).

Experimental Part

General. RP-18 Silica gel (SiO₂, 40–65 μm): Merck Company, Germany. Sephadex LH-20 (40–70 μm): Amersham Pharmacia Biotech AB, Uppsala, Sweden. Silica gel (SiO₂, 200–300 mesh) for column chromatography (CC) and GF₂₅₄ for TLC: Qindao Marine Chemical Factory, Qingdao, P. R. China. Compounds were detected under UV (254 nm and 365 nm), and after spraying with an anisaldehyde sulfuric acid soln., followed by heating. Optical rotation: Horiba SEAP-300 spectropolarimeter. UV Spectra: Shimadzu double-beam 210A spectrophotometer. IR Spectra: Bruker Tensor 27 spectrometer with KBr pellets. 1D- and 2D-NMR Spectra: Bruker DRX-500 MHz NMR spectrometer with TMS as internal standard. Pos. ESI-MS and HR-ESI-MS Spectra: VG Autospec-3000 spectrometer.

Plant Material. The aerial parts of *B. asiatica* LOUR were collected in Guangxi Province, P. R. China, in July, 2006, and identified by Mrs. Chun-Xia Zeng, Kunming Institute of Botany, Chinese Academy of Sciences. A voucher specimen (Zeng 0718) has been deposited with the Herbarium of Kunming Institute of Botany, Chinese Academy of Sciences.

Extraction and Isolation. The air-dried aerial parts (12.5 kg) were crushed and extracted with 90% aq. MeOH (40 l × 4) at r.t. (48 h × 4). After evaporation of the solvent under reduced pressure, the viscous residue was redissolved in H₂O, and then partitioned with AcOEt (15 l × 4) to afford AcOEt and H₂O layers. The AcOEt fraction (218 g) was chromatographed on a SiO₂ column (2.0 kg, 200–300 mesh), using a CHCl₃/acetone gradient (from CHCl₃ to CHCl₃/acetone 1:1), to give 9 fractions (I–IX). Fr. VII (21.0 g) was submitted to SiO₂ CC (600 g) and eluted with petroleum ether (PE)/acetone 8:1, 6:1, 4:1, 2:1, 1:1, to yield five subfractions (A–E). Subfrs. B and E were purified by recrystallization from cold MeOH to give *O*-methylferulic acid (9.2 mg) and *p*-methoxycinnamic acid (30 mg), respectively. Subfr. C was purified by RP-18 CC with MeOH/H₂O 80:20, 85:15, 90:10 to afford compounds **4** (208.5 mg) and **3** (86.6 mg). Fr. VIII (15.0 g) was submitted to SiO₂ CC (450 g) and eluted with PE/AcOEt 6:1, 4:1, 2:1, to yield three subfractions (F–H). Subfr. G was first applied to RP-18 CC with MeOH/H₂O 60:40, 70:30, 75:25, 80:20, and then to Sephadex LH-20 (MeOH) to give compounds **5** (834.2 mg), **2** (57.8 mg), and **1** (88.5 mg). Subfr. H was purified by recrystallization in cold MeOH to give ferulic acid (9.8 mg).

Asiatiside A (=6-Deoxy-4-O-[(2E)-3-(4-methoxyphenyl)-1-oxoprop-2-en-1-yl]-α-L-mannopyranose 3-Acetate; **1**). White amorphous powder. $[\alpha]_D^{25} = +85.7$ ($c = 0.21$, acetone). UV (MeOH): 311 (3.82), 298 (3.28), 222 (2.21), 208 (2.66). IR (KBr): 3463, 3385, 2972, 1738, 1692, 1629, 1610, 1512, 1040. ¹H- and ¹³C-NMR: Tables 1 and 2, resp. ESI-MS (pos.): 755 ([2 M + Na]⁺), 389 ([M + Na]⁺). HR-ESI-MS: 389.1211 ([M + Na]⁺, C₁₈H₂₂NaO₈⁺; calc. 389.1212).

Asiatiside B (=6-Deoxy-4-O-[(2E)-3-(4-hydroxy-3-methoxyphenyl)-1-oxoprop-2-en-1-yl]-α-L-mannopyranose 3-Acetate; **2**). White amorphous powder. $[\alpha]_D^{25} = +56.1$ ($c = 0.19$, acetone). UV (MeOH): 327 (1.34), 298 (1.16), 244 (0.95), 205 (2.43). IR (KBr): 3422, 3376, 2941, 1737, 1692, 1629, 1610, 1512, 1041. ¹H- and ¹³C-NMR: Tables 1 and 2, resp. ESI-MS (pos.): 787 ([2 M + Na]⁺), 764 ([2 M]⁺), 405 ([M + Na]⁺). HR-ESI-MS: 405.1161 ([M + Na]⁺, C₁₈H₂₂NaO₉⁺; calc. 405.1161).

Asiatiside C (=6-Deoxy-4-O-[(2E)-3-(3,4-dimethoxyphenyl)-1-oxoprop-2-en-1-yl]-α-L-mannopyranose 2-Acetate; **3**). White amorphous powder. $[\alpha]_D^{25} = +115.2$ ($c = 0.22$, acetone). UV (MeOH): 323 (1.39), 297 (1.09), 235 (0.95), 205 (2.28). IR (KBr): 3445, 3316, 2940, 1741, 1704, 1633, 1597, 1517, 1067. ¹H- and ¹³C-NMR: Tables 1 and 2, resp. ESI-MS (pos.): 815 ([2 M + Na]⁺), 419 ([M + Na]⁺), 397 ([M + H]⁺). HR-ESI-MS: 419.1319 ([M + Na]⁺, C₁₉H₂₄NaO₉⁺; calc. 419.1318).

Asiatiside D (=6-Deoxy-4-O-[(2E)-3-(4-methoxyphenyl)-1-oxoprop-2-en-1-yl]-α-L-mannopyranose 2-Acetate; **4**). White amorphous powder. $[\alpha]_D^{25} = +58.7$ ($c = 0.24$, DMSO). UV (MeOH): 309 (3.84), 298 (3.40), 222 (2.36), 208 (2.71). IR (KBr): 3463, 3385, 2962, 1741, 1704, 1629, 1597, 1517, 1067. ¹H- and ¹³C-NMR: Tables 1 and 2, resp. ESI-MS (pos.): 755 ([2 M + Na]⁺), 389 ([M + Na]⁺), 367 ([M + H]⁺). HR-ESI-MS: 389.1203 ([M + Na]⁺, C₁₈H₂₂NaO₈⁺; calc. 389.1212).

Known Compounds. The structures of the known compounds were identified as buergeriside C₁ (4-*O*-(*p*-methoxycinnamoyl)-α-L-rhamnopyranose; **5**) [18], *p*-methoxycinnamic acid [14a], ferulic acid [15][19], *O*-methylferulic acid [15] by ESI-MS, and ¹H- and ¹³C-NMR spectral data.

Acid Hydrolysis of 1–4. Compounds **1–4** (20 mg each) were refluxed with 10% HCl/MeOH (20 ml) on a H₂O bath at 60° for 5 h. After cooling, the mixture was evaporated to dryness and redissolved in

H₂O, then partitioned with AcOEt, to afford AcOEt and H₂O layers. The sugar was identified as rhamnose by TLC comparison with an authentic sample using CHCl₃/MeOH 6 : 4, $R_f = 0.62$. Purification of the aq. layer was performed by prep. TLC eluted four times with CHCl₃/MeOH/H₂O 70 : 30 : 1 to afford L-rhamnose ($R_f = 0.68$) with positive values of $[\alpha]_D$ (11.1, 11.8, 12.2, and 13.6 from **1–4**, resp.). The AcOEt soln. was monitored by HP-TLC SiO₂ GF₂₅₄ plate using CHCl₃/acetone 5 : 1 and PE/acetone 2 : 1, and showed several decomposition products.

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